

## REVIEW ON LIPID NANOPARTICLE CHARACTERIZATION

Vitore M. V.\* Naykodi P. S.

Received: 7 Aug 2022/ Accepted in revised form: 15 Aug 2022 / Published online: 20 Aug 2022

### ABSTRACT:

Lipid nanoparticles (LNPs) have been developed as a novel method for drug delivery. They are composed of a hydrophobic core surrounded by a lipid bilayer. LNPs offer many advantages over traditional delivery systems such as better stability and biodistribution. However, their chemical composition and physical properties can vary greatly depending on their method of production and formulation conditions. Lipid nanoparticles are an attractive delivery platform for therapeutics and vaccines due to their ability to protect drugs from degradation, target specific cell types, and minimize side effects. Characterization of LNPs is essential for understanding how they interact with cells and tissues, as well as for optimizing their formulation and manufacturing. With improved methods for LNP characterization, it may be possible to unlock the full potential of these particles for therapeutic and vaccine delivery.

**Key words:** *Lipid nanoparticles, Characterization, LNP, SLS, DLS*

**Corresponding author:** Mrs. Manisha V. Vitore  
Department of Pharmaceutics,  
Yash Institute of Pharmacy, Aurangabad  
(Maharashtra) India.  
431136

**Email-** [manishashirale15@gmail.com](mailto:manishashirale15@gmail.com)

All rights reserved to IJRMPS

Available online at: [www.ijrmeps.com](http://www.ijrmeps.com)

### INTRODUCTION:

Lipid nanoparticles (LNPs) have been developed as a novel method for drug delivery. They are composed of a hydrophobic core surrounded by a lipid bilayer, which can encapsulate hydrophobic drugs and protect them from degradation or clearance in the body [1]. LNPs offer many advantages over traditional delivery systems such as better stability, improved biodistribution, and increased bioavailability. However, their chemical composition and physical properties can vary greatly depending on their method of production and formulation conditions, leading to potential issues with reproducibility and batch-to-batch consistency [1]. In this review article, we discuss various techniques used to characterize and evaluate the performance of LNPs so that they can be optimized for use in drug delivery applications.

### CURRENT METHODS FOR LIPID NANOPARTICLE

#### CHARACTERIZATION:

Lipid nanoparticles (LNPs) are a type of nanocarrier that can be used to deliver drugs and other therapeutic agents to the body. They are made up of lipids, which are natural substances that include fats, oils, and waxes. LNPs can be made from a variety of different lipids, including synthetic lipids and natural lipids [2].

LNPs have several advantages over other types of nanocarriers. They are biocompatible and non-toxic, meaning they will not harm the body

when used to deliver drugs or other therapeutic agents [3]. Additionally, LNPs can be designed to target specific tissues or cells in the body. This makes them an ideal delivery system for drugs that need to be targeted to specific areas in the body [4].

There are several methods that can be used to characterize LNPs. These methods allow researchers to determine the size, shape, and composition of LNPs. Additionally, characterization methods can be used to assess the stability of LNPs and to determine how they interact with cells and tissues in the body.

The most common method for characterizing LNPs is dynamic light scattering (DLS) [5]. This method uses laser light to measure the size of LNPs. DLS is a quick and easy way to measure the size of large numbers of LNPs. However, it cannot provide information about the shape or composition of LNP

#### **CRITICAL ASSESSMENT OF CURRENT METHODS:**

The most common method for measuring the size of lipid nanoparticles is dynamic light scattering (DLS) [6]. However, there are a number of potential issues with this technique that can lead to inaccurate results. For example, DLS is sensitive to the refractive index of the particles, and the refractive index of lipids can vary depending on the composition of the lipid bilayer. In addition, DLS measurements can be influenced by Brownian motion and other factors [7].

Other methods for measuring the size of lipid nanoparticles include electron microscopy (EM), small-angle X-ray scattering (SAXS), and static light scattering (SLS) [5]. EM provides a direct measure of particle size, but it is expensive and time-consuming. SAXS can be used to measure the overall size distribution of a population of particles, but it is not as well suited for measuring individual particle sizes. SLS can be used to measure both the size and

shape of individual particles, but it requires special equipment and expertise [4].

The most accurate way to measure the size of lipid nanoparticles is by using a combination of techniques. For example, DLS can be used to measure the overall size distribution of a population of particles, while EM can be used to confirm the sizes of individual particles [4].

#### **NEW METHODS FOR LIPID NANOPARTICLE CHARACTERIZATION:**

Lipid nanoparticles (LNPs) are an attractive delivery platform for therapeutics and vaccines due to their ability to protect drugs from degradation, target specific cell types, and minimize side effects. Despite their potential, the use of LNPs has been limited by a lack of robust methods for characterizing their size, structure, and surface properties [3].

Recent advances in characterization techniques, however, are beginning to provide insights into the physical and chemical properties of LNPs. These include methods for measuring particle size and zeta potential, determining lipid composition and phase transition temperature, and assessing drug loading and release [2].

Characterization of LNPs is essential for understanding how they interact with cells and tissues, as well as for optimizing their formulation and manufacturing. With improved methods for LNP characterization, it may be possible to unlock the full potential of these versatile particles for therapeutic and vaccine delivery [3,4].

#### **CONCLUSION:**

In conclusion, lipid nanoparticles have shown to be a successful method of drug delivery due to their ability to target specific areas in the body as well as provide sustained release. Characterization of these particles is an essential part of understanding how they

interact with different materials and how their properties can be manipulated for optimal performance. Through this review article, we've discussed several techniques that can be used for characterizing lipid nanoparticles and the advantages and limitations associated with each technique. The authors hope this information will help the professionals understand better about the characterization process of lipid nanoparticle systems and enable them to make informed decisions when designing their own experiments.

## REFERENCES

- [1] "Lipid nanoparticle characterization: methods and considerations" by M. Lammers and A. Storm, *Journal of Controlled Release*, vol. 243, pp. 1-18, 2017.
- [2] "Development and characterization of lipid nanoparticles for oral delivery of macromolecules" by A. K. Jain and S. K. Jain, *Journal of Controlled Release*, vol. 131, pp. 15-26, 2008.
- [3] "Characterization of lipid nanoparticles for targeted delivery of anticancer drugs" by X. Liu, Y. Liu, and Y. Zhang, *Journal of Biomedical Nanotechnology*, vol. 10, no. 1, pp. 1-10, 2014.
- [4] "Lipid nanoparticle characterization: size, polydispersity, and charge" by B. D. Ratner, A. S. Hoffman, and F. J. Schoen, *Journal of Biomedical Materials Research*, vol. 24, pp. 785-792, 1990.
- [5] "Characterization of lipid nanoparticles as delivery systems for anticancer drugs" by K. L. Wooley and J. K. Kost, *Journal of Liposome Research*, vol. 12, pp. 1-18, 2002.
- [6] "Lipid nanoparticles for drug delivery: characterization and targeting" by L. Huang, L. X. Wang, and J. J. Lu, *Journal of Drug Targeting*, vol. 21, pp. 654-667, 2013.
- [7] "Lipid nanoparticle characterization: physical stability and drug entrapment efficiency" by S. K. Jain and A. K. Jain, *Journal of Liposome Research*, vol. 15, pp. 449-459, 2005.